

Performance Quality Validation of Autoclave Vacuum, Liquid and Gravity Displacement Cycles for Sterilization of Biohazardous Waste

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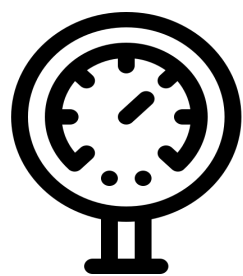
INTRODUCTION

- The development of a standardized autoclave validation procedure guarantees the reliable sterilization of complex biohazardous waste (BHW) generated in public health laboratories in compliance with international biosafety standards.
- This study assesses the precise functions, strengths, and limitations of autoclave pre-vacuum, liquid, and gravity displacement cycles, validating appropriate cycles for routine sterilization.
- Results support the optimization of cycle parameters and model waste loads to achieve reliable sterilization across a diverse range of wastes and cycles.

Objectives

- Design representative waste models to reflect real BHW across multiple labs.
- Simulate default lab protocols with model waste and collect data.
- Identify factors impacting reliability: Operational quality, waste contents, volume, time, heat and steam penetration.
- Recommend evidence-based process improvements aligned with established protocols.
- Standardize validation procedures for future use.

Autoclave Quality Controls



Operational Quality (OQ)

- Reliable steam/pressure; Routine QA maintenance.
- Monitoring with chemical integrators and steam/pressure quality checks.



Performance Quality (PQ)

- Reliable sterilization of biohazardous waste.
- Sterility assurance level (SAL) of 10^{-6} (1.0 CFU / 1,000,000 cycles).
- Compliant with regulatory benchmarks.

Bearss et al. 2017

Table 1. Autoclave Cycle Types

Cycle Type	Purpose	Sterilization	Feature
Liquid 	General-Purpose, Liquid media.	Slow increase in steam, heat and pressure to prevent boiling.	Slow Exhaust Slow pressure release to prevent flash boiling.
Gravity 	General-Purpose, Reusable instruments.	High-velocity steam injections force chamber air out the drain.	Drying Chamber air evacuated for a set time.
Vacuum 	Dry / wrapped waste; Sensitive instruments.	Strong vacuum pulses remove air, followed by steam pressure injections.	Drying Chamber air evacuated for a set time.

Table 2. SHL DCD Autoclave Models

Autoclave Model	Brand	Cycle Options	Vacuum Capability
N936675-01 X1	CSS	<ul style="list-style-type: none">VacuumGravityLiquid	Venturi + Liquid-Ring Pump (≤ 30 inHg)
SSR-3A-ADVPRO	CSS	<ul style="list-style-type: none">GravityLiquid	Venturi (10-15inHg)
46400D/2140DD	Easter Services	<ul style="list-style-type: none">Liquid	N/A

METHODS

Designing Representative Waste

- Real biohazardous waste (BHW) cannot safely be tested for sterility. **Representative waste** is designed to simulate BHW.
- Reliable sterilization depends on accurately predicting waste composition, which varies by laboratory type.
- Lab experts are surveyed to assist with designing accurate models for validation.

Le et al. (2005)

Table 3. Biohazard Waste Compositions

Laboratory Response Network (LRN)		
LRN-M1 (Standard Waste)	LRN-M2 (BSL-3 PPE)	
<ul style="list-style-type: none">4lbsSharps containers were buried beneath dry PPE and cloth	<ul style="list-style-type: none">2lbsGownsGlovesAnkle guardsFace shields	
Reference Bacteriology (RB)		
RB-M1 (Primary Benchmark Waste)	RB-M2 (BSC Waste)	RB-M3 (BSC Sharps)
<ul style="list-style-type: none">16lbs250 agar plates	<ul style="list-style-type: none">9lbs120 agar plates	<ul style="list-style-type: none">1lb

Model Bag and Chamber Configuration

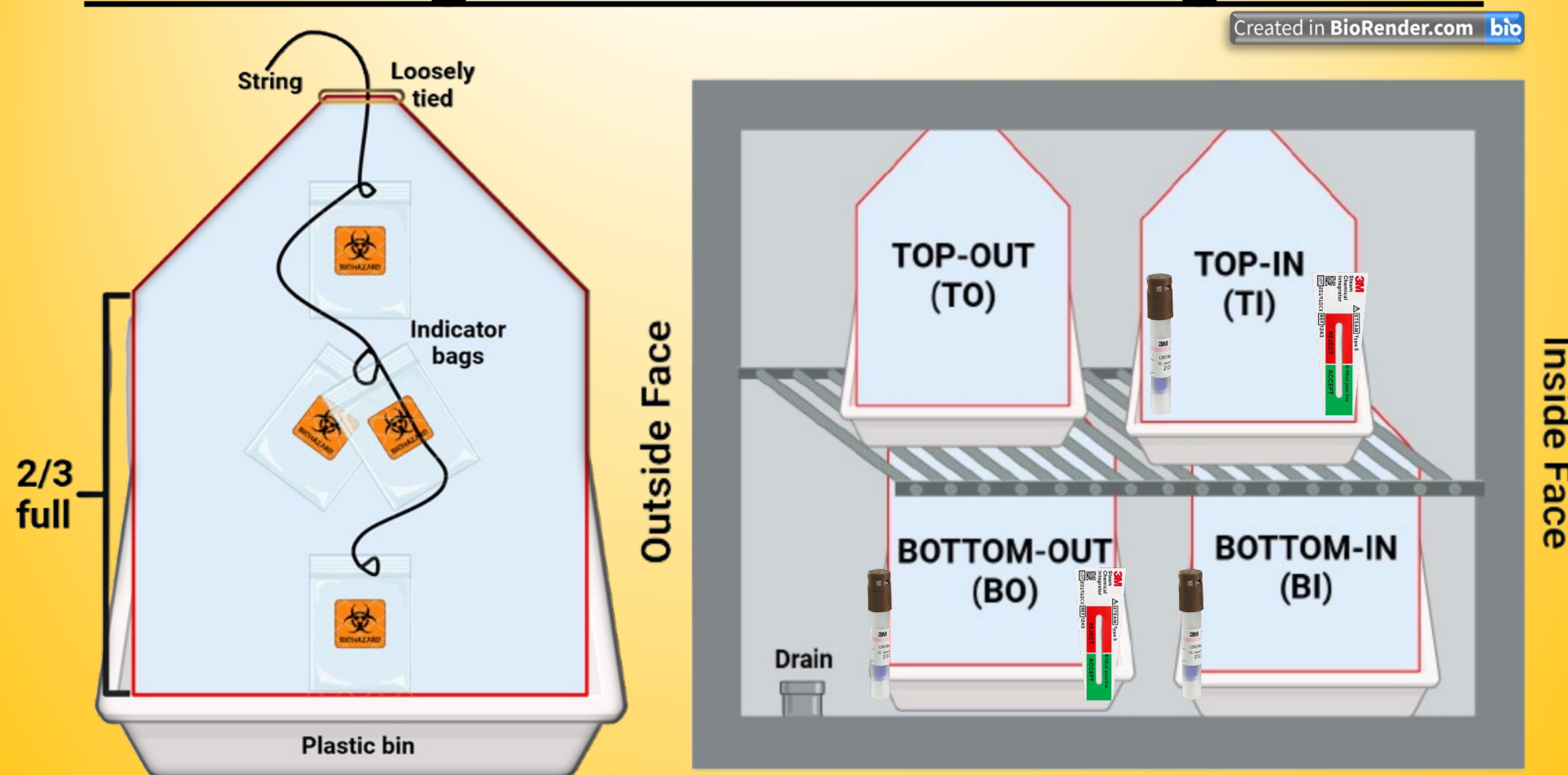


Figure 1. Indicator Bag Arrangement and Chamber Load Labelling
Technical specifications between bag and autoclave models vary between laboratories. Bags were loosely tied pre-sterilization to facilitate steam penetration.

Table 4. Sterilization Indicators

Indicator Type	Methods & Features	Significance
Chemical (CI) 	<ul style="list-style-type: none">Molten pellet migrates past the accept line.Mimics BI response but non-confirmatory.	<ul style="list-style-type: none">First validation step.Rapid assessment of sterility before BI confirmation.
Biological (BI) 	<ul style="list-style-type: none">Fluorescently tagged, heat-resistant bacterial spores (10^6 CFU/mL).Auto-reader detects growth in 3 hours; pH/color change confirms after 48 hours.	<ul style="list-style-type: none">Confirmatory IndicatorValidates sterilization to a Sterility Assurance Level (SAL) of 10^{-6}. Required for final validation.

RESULTS

Triplicate Validation Standard

- All indicators in a cycle must pass.
- A cycle setting and load model must be successfully run in triplicate to be validated.
- A cycle abortion or indicator failure during the triplicate resets the validation.
- Successful triplication confirms PQ validation for designated cycle setting and bag model.
- All cycles are documented for compliance tracking.

Bearss et al. 2017

Sterilization Time by Bag Model and Cycle Type

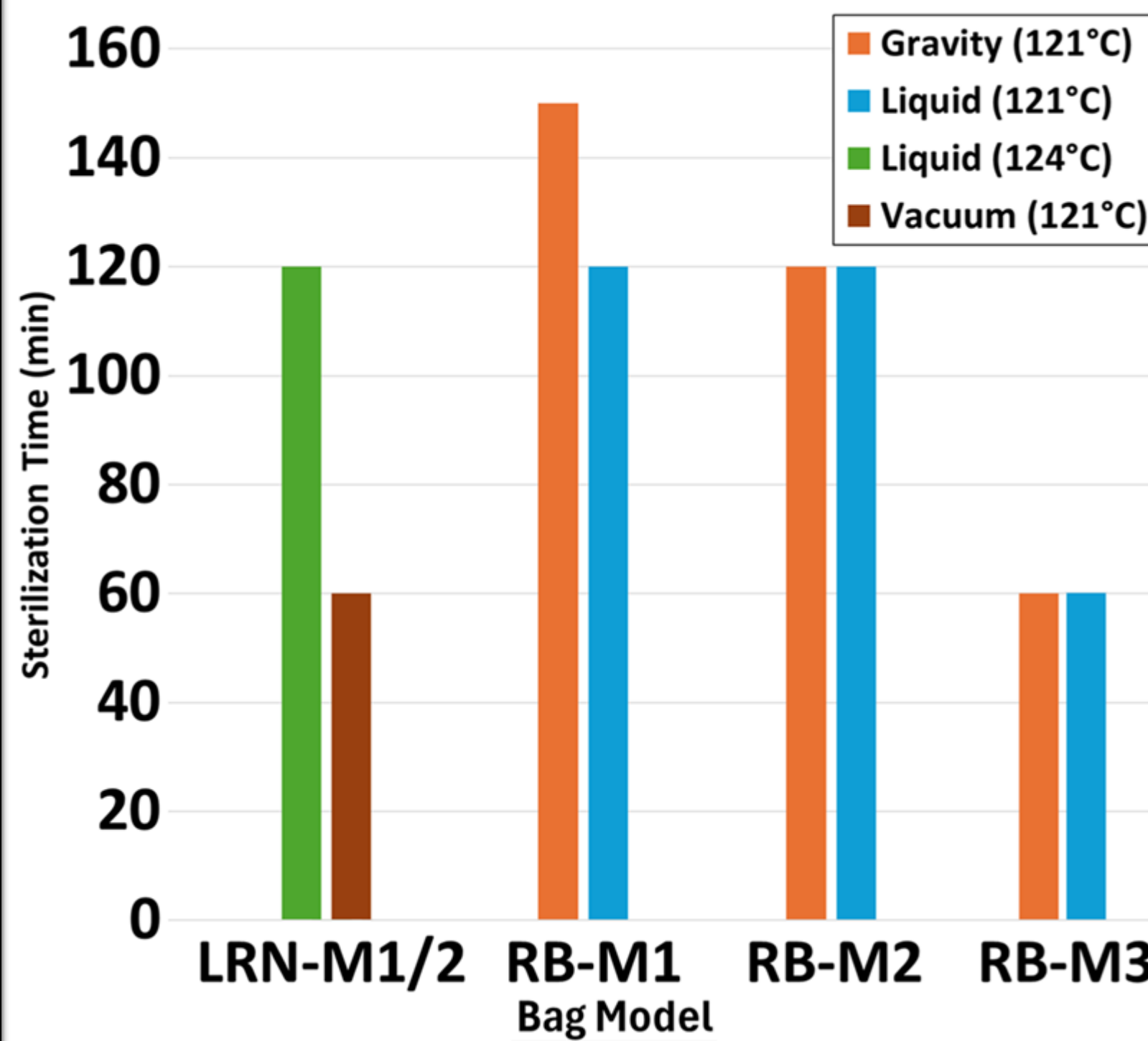


Figure 2. Validated Sterilization Times by Bag Model and Cycle Type
*LRN-M1/2, Liquid 124C used 500mL H2O in bags

Optimizing Sterilization Time to Waste Mass

- 121°C Liquid cycle mass threshold tests were performed with modified RB-M1 bags containing 250 (16lbs), 120 (9lbs), and 50 (3lbs) agar plates. Agar plates are the primary waste mass in these loads.
- 60 minutes failed to sterilize 50-plate bags.
- 90 minutes for 50-plate bags is still under evaluation:
 - Preliminary results show potential failures.
- 120 minutes was required for both 120-plate (9lbs) and 250-plate (16lbs) bag sterilization.
- Data from 120-minute tests may suggest a plateau in time-per-weight optimization.
 - Bearss et al. (2017) proposes that chamber volume—not load mass—is the true upper limit to sterilization capacity.
- A 150-minute Gravity cycle was validated for 250-plate bags, compared to the validated 120-minute Gravity cycle for the 120-plate bags—potentially attributed with the liquid cycle “slow-exhaust” phase—demonstrating the need for extended times with less efficient cycles.

Significance of The Drying Cycle Drying Cycles Are Not Part of Sterilization

- Post-sterilization, in Vacuum and Gravity cycles, the vacuum evaporates condensation in the chamber.
- During drying, the chamber rapidly cools below sterilization conditions (e.g., $<121^{\circ}\text{C}$, <15 psi). No new steam is injected.
- Useful for handling and packaging—but not sterilization.

DISCUSSION

Extreme Challenge Conditions

CHALLENGES

- Steam Penetration**
 - Sharps containers.
 - Densely-packed dry waste.
 - Sealed bags within bags.

Bearss et al. (2017)

Heat Distribution

- Liquid/Gravity cycles need more time to sterilize dry lab waste.
- Waste microclimates inhibit even heating.



Figure 3. Sharps Container Microclimate Challenge
Sealed containers trap cool air, preventing steam penetration and sterilization.

AUTOClave PROCESS IMPROVEMENTS

- Vacuum cycles for high-challenge waste with minimal water content to remove cool-air pockets.
- Avoid excessive layering and seals (triple or quadruple layers) prior to sterilization.
- To reduce sterilization time, autoclave sharps separately.
- Adding water to bags facilitates steam penetration for liquid and gravity cycles.

Le et al. (2005)

Validation Considerations

- Verify Operational Quality prior to PQ testing.
- Prioritize threshold-volume validations for efficient use of autoclave space and power.
- Use failures to identify steam penetration issues. Adjust only one variable at a time to isolate impediments.
- Suggest evidence-based process improvements:
 - Separation of sharps waste.
 - Excessive bagging or pre-sealing.
 - Alternative cycle parameters.
- Significant autoclave repairs, configuration changes, or failed PQs will reset validation.

Future Directions

- Extend PQ validation procedure to all SHL DCD autoclave models with model waste.
- Collect evidence to drive process improvements where needed.
- Publication of an official validation procedure in the SHL Quality and Operations Records.
- Use of data loggers to measure precise conditions in model bags.

References Acknowledgements

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